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Total synthesis of sporochnols, fish deterrents from a marine alga

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Abstract—Sporochnols, fish deterrents from a marine alga, were synthesized, using intramolecular CH insertion of alkylidenecarbene as a key step to construct the chiral quaternary center. © 2002 Elsevier Science Ltd. All rights reserved.

Sporochnols (**1**–**3**) are monoterpene-substituted phenols isolated from the marine alga *Sporochnus bolleanus*. 1 They are known to be chemical defense substances and to exhibit feeding deterrence activity toward reef fishes. Configuration of the asymmetric center of (+)-sporochnol A (**1**) was determined as *S* by the enantioselective synthesis of (−)-ent-1 by Ogasawara's group in 1997.² Each sporochnol B (**2**) and C (**3**) which includes two stereocenters in one molecule is a 1:1 diastereomeric mixture. Syntheses of racemic or optically active sporochnol A (**1**) have recently been reported by several groups.³ We describe here the synthesis of (\pm) -1 and both enantiomers of 1 , using the $C-H$ insertion reaction of alkylidenecarbene as the key step.⁴ $(+)$ -1 was also converted into **2** and **3** enriched with one of the diastereomers to make assignment of NMR spectra of all the diastereomers possible.

Alkylation of the enolate of the ester **4** with iodomethane in the presence of DMPU in THF gave (\pm) -5 in good yield. Reduction of ester (\pm) -5 with lithium aluminum hydride gave alcohol (\pm) -6, which was oxidized under Swern's conditions followed by Wittig reaction with 1-triphenylphosphoranylidene-2 propanone to afford α , β -unsaturated ketone (\pm)-7. Hydrogenation of **7** in the presence of palladium on charcoal gave methyl ketone (\pm) -8. Exposure of (\pm) -8 with lithiotrimethylsilyldiazomethane⁵ produced cyclopentene (\pm) -9 cleanly via intramolecular C-H insertion of the alkylidenecarbene. It is known that the reaction proceeds with a retention of configuration.⁶ The high yield (92%) of this reaction is attributable to the low dissociation energy of the benzylic $C-H$ bond and/or the electron donating effect of *p*-methoxy group.

Cyclopentene (\pm) -9 was subjected to epoxidation with *m*-chloroperbenzoic acid to give epoxide (\pm) -10 as a diastereomeric mixture. The reaction of (±)-**10** with diethylaluminum *N*,*N*-tetramethylpiperazide⁷ at room temperature gave the rearranged allyl alcohol (\pm) -11. The possible isomer, which contains an *endo* double bond in the five-membered ring, was not produced at all. Dihydroxylation of the double bond of (\pm) -11 using OsO₄ under catalytic conditions gave triol (\pm) -12 as a diastereomeric mixture. The stereochemistry of (±)-**10** and (\pm) -12 was not examined in detail because stereocenters except the benzylic quaternary center would disappear in the next step. Cleavage of two $C-C$ bonds of (\pm) -12 occurred by treatment with excess NaIO₄ via the intermediate (\pm) -13 to give aldehyde (\pm) -14. Direct reaction of (\pm) -14 with excess methylenetriphenylphosphorane in DMSO gave the desired product (\pm) -15 in good yield. Reduction of acid (±)-**15** with lithium alu- * Corresponding author. minum hydride in THF gave alcohol (±)-**16**. The con-

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version of (\pm) -12 to (\pm) -16 was achieved without chromatographic separation of intermediates in 65% overall yield. Swern oxidation of alcohol (±)-**16**, followed by the Wittig reaction using isopropyridenetriphenylphosophorane gave diene (±)-**17**. Although the reaction of (\pm) -17 and boron tribromide gave no desired product due to the addition of generated hydrogen bromide to double bonds, treatment of (\pm) -17 with Ph_2PLi^8 at 50°C afforded the (\pm) -1 in good yield. ¹H NMR, 13C NMR, IR, and MS spectra of the synthetic compound were essentially identical to those of the natural product.¹

Establishing the pathway from ester 4 to (\pm) -1, our attention then turned to the preparation of optically active sporochnols. Since Matsumoto et al. reported the effective enzymatic resolution of 1-acetoxy-2- (aryl) propanes, \degree we applied their conditions to acetate (\pm) -18, derived from (\pm) -6. (\pm) -18 was hydrolyzed with PPL in methanol to give acetate (−)-**18** and alcohol (−)-6 ([α]²⁵ −16.8 (*c* 1.25, CHCl₃) in 49% and 51% yield, respectively (Scheme 1). Hydrolysis of (+)-**18** with aqueous KOH in methanol gave $(+)$ -6 ($[\alpha]_D^{25}$ +16.8 (*c* 1.37, $CHCl₃$) (Scheme 2). The absolute configuration of the alcohol (6) was known¹⁰ and the enantiomeric excess of (−)-**6** and (+)-**6** was calculated from NMR spectra of MTPA esters of the alcohols as 93% ee and 95% ee, respectively. *E* value of the kinetic resolution was 96.4.

In the same way as the synthesis of (\pm) -1, $(+)$ -6 was converted to (−)-**7** and then into (+)-**8**. Although the small value of the optical rotations of (−)-**7** (−1.3°) and (+)-**8** (1.5°) implied occurrence of partial racemization of the intermediate, (+)-**8** was converted into the final compound **1**. The synthetic sporochnol A thus obtained showed essentially no optical rotation. Racemization of the aldehyde during the Wittig reaction was significant,

Scheme 1. *Reagent and conditions*: (i) LDA; MeI, DMPU, THF, −78°C (quant.); (ii) LiAlH₄, Et₂O, rt (98%); (iii) DMSO (COCl)₂, Et₃N, CH₂Cl₂, −78°C; (iv) Ph₃P=CHCOCH₃, benzene, 60°C (67% from 6); (v) H₂, EtOAc, Pd/C, rt (89%); (vi) TMSC(Li)N₂, THF, 0°C (92%); (vii) *m*CPBA, NaHCO₃, CH₂Cl₂, rt (91%); (viii) LTMP, Et₂AlCl, toluene, 0°C (quant.); (ix) OsO₄, NMO, THF-H₂O, rt (96%); (x) NaIO₄, THF, H₂O, rt; (xi) Ph₃P=CH₂, DMSO, 40°C; (xii) LiAlH₄, (65% from **12**); (xiii) DMSO, (COCl)₂, Et3N, CH2Cl2, −78°C; (xiv) Ph3P-C(CH3)2, DMSO, 40°C (74% from **16**); (xv) Ph2PLi, THF, 50°C (85%).

Scheme 2. *Reagent and conditions:* (i) PPL, MeOH, buffer, 35°C (quant.); (ii) KOH, MeOH, H₂O, rt (97%).

so that modification of the route which excludes the aldehyde as an intermediate is imperative.

After many trials, we found that the tosylate **21** obtained from (+)-**6** reacted with sodium cyanide cleanly to give nitrile $(+)$ -22 ($[\alpha]_D^{25}$ +8 (*c* 0.19, CHCl₃)) (Scheme 3). (+)-**22** was reduced with DIBAL to the aldehyde that was submitted to Wordsworth–Emmons reaction using triethyl phosphonoacetate and butyllithium, giving α,β-unsaturated ester 23 as a *cis–trans* mixture. After hydrolysis of the ester, the carboxylic acid was treated with DPPA and triethylamine under Shioiri's conditions.¹¹ The Curtius rearrangement product thus obtained was hydrolyzed without isolation to give methyl ketone $(+)$ -8 $([\alpha]_D^{25}$ +17 (*c* 0.29, CHCl₃). (+)-8 was transformed to (+)-1 ($[\alpha]_D^{25}$ +5.8 (*c* 0.95, CHCl₃); lit.^{3a}: [α]_D³⁰ +2.9 (*c* 1, CHCl₃); natural¹: $[\alpha]_D$ +10 (*c* 1, CHCl₃)), using the established sequence as shown in Scheme 1. The physical properties of synthetic sporochnol A were identical to those of the

natural product. (−)-1 ($[\alpha]_D^{25}$ –5.4 (*c* 1.48, CHCl₃); lit.^{3b}: $[\alpha]_{D}^{\overline{20}}$ –2.5 (*c* 1, CHCl₃)) was also obtained in the same way from (−)-6. As the yield of each step was good, we were able to obtain about 0.5 g of both enantiomers of sporochnol A.

Conversions of (+)-**1** into other sporochnols were achieved as follows (Scheme 4). Epoxidation of (+)-**1** with *m*-chloroperbenzoic acid gave **2** as a 1:1 diastereomeric mixture. Treatment of **2** with diethylaluminum *N*,*N*-tetramethylpiperazide afforded **3** in good yield. ¹ H NMR and 13C NMR spectra of **2** and **3** were identical with those of natural products. For the complete assignment of the NMR spectra of **2** and **3**, one of the diastereomers of **2** (**2a**, 62% de) was prepared in three steps (protection, Shi asymmetric epoxidation12 and deprotection). Rearrangement of **2a** afforded $3a$, thus all signals of ¹H NMR and ¹³C NMR of **2** and **3** were assigned to each diastereomer.¹³

Scheme 3. *Reagent and conditions:* (i) TsCl, Py, CH₂Cl₂, rt (93%); (ii) NaCN, DMSO, 80°C (89%); (iii) DIBAL, toluene, rt; (iv) (EtO)₂P(O)CH(Me)COOEt, BuLi, THF, rt (93% from 22); (v) KOH, H₂O, MeOH, rt; (vi) DPPA, Et₃N, toluene, rt; (vii) HCl, H2O (62% from **23**).

Scheme 4. *Reagent and conditions:* (i) *mCPBA*, NaHCO₃, CH₂Cl₂, rt (quant.); (ii) LTMP, Et₂AlCl, toluene, 0°C (96%); (iii) TBDMSOTf, 2,6-di-tert-butyl-4-methylpyridine, CH₂Cl₂, 0°C (95%); (iv) **24**, oxone, *n*-Bu₄NHSO₄, buffer, dimethoxymethane, rt (65%); (v) TBAF, THF, 0°C (98%).

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- 13. **2a**: ¹H NMR (500 MHz, CDCl₃) δ 7.166 (2H, d, *J*=8.7 Hz), 6.769 (2H, d, *J*=8.7 Hz), 5.987 (1H, dd, *J*=10.7, 17.4 Hz), 5.086 (1H, dd, *J*=10.7, 1.2 Hz), 5.035 (1H, dd, *J*=17.4, 1.2 Hz), 2.691 (1H, t, *J*=6.3 Hz), 1.909 (1H, dt, *J*=12.8, 4.6 Hz), 1.805 (1H, dt, *J*=12.8, 4.9 Hz), 1.24– 1.50 (2H, m), 1.347 (3H, s), 1.284 (3H, s), 1.170 (3H, s); ¹³C NMR (125 MHz, CDCl₃) δ 153.66, 146.81, 138.98, 127.77, 114.88, 111.73, 64.81, 58.70, 43.39, 37.46, 24.87, 24.83, 24.21, 18.61. **2b** (diastereomer of **2a**): ¹ H NMR $(500 \text{ MHz}, \text{CDCl}_3) \delta$ 7.166 (2H, d, $J=8.7 \text{ Hz}$), 6.769 (2H, d, *J*=8.7 Hz), 5.980 (1H, dd, *J*=10.7, 17.4 Hz), 5.105 (1H, dd, *J*=10.7, 1.2 Hz), 5.045 (1H, dd, *J*=17.4, 1.2 Hz), 2.694 (1H, t, *J*=6.3 Hz), 1.994 (1H, ddd, *J*=13.4, 9.6, 7.6 Hz), 1.723 (1H, ddd, *J*=13.4, 9.4, 7.4 Hz), 1.24–1.50 (2H, m), 1.342 (3H, s), 1.293 (3H, s), 1.206 (3H, s); ¹³C NMR (125 MHz, CDCl₃) δ 153.68, 146.56, 139.15, 127.69, 114.90, 111.96, 64.81, 58.70, 43.37, 37.38, 24.87, 24.98, 24.25, 18.56. **3a**: ¹H NMR (500 MHz, CDCl₃) δ 7.147 (2H, d, *J*=8.3 Hz), 6.746 (2H, d, *J*=8.3 Hz), 5.984 (1H, dd, *J*=10.7, 6.7 Hz), 5.070 (1H, d, *J*=12.2 Hz), 5.012 (1H, d, *J*=17.7 Hz), 4.915 (1H, bs), 4.834 (1H, bs), 4.012 (1H, t, *J*=6.4 Hz), 1.764 (1H, dt, *J*=13.0, 4.9 Hz), 1.670 (1H, dt, *J*=13.0, 4.9 Hz), 1.34–1.51 (2H, m), 1.621 (3H, s), 1.329 (3H, s); ¹³C NMR (125 MHz, CDCl₃) δ 153.70, 147.03, 147.02, 139.13, 127.74, 114.91 111.60, 111.50, 76.58, 43.26, 36.64, 29.60, 25.08, 17.35. **3b** (diastereomer of **3a**): ¹H NMR (500 MHz, CDCl₃) δ 7.147 (2H, d, *J*=8.3 Hz), 6.746 (2H, d, *J*=8.3 Hz), 5.977 (1H, dd, *J*=10.7, 6.7 Hz), 5.070 (1H, d, *J*=12.2 Hz), 5.012 (1H, d, *J*=17.7 Hz), 4.915 (1H, bs), 4.834 (1H, bs), 4.012 (1H, t, *J*=6.4 Hz), 1.853 (1H, dt, *J*=12.9, 4.6 Hz), 1.587 (1H, dt, *J*=12.9, 4.6 Hz), 1.34–1.51 (2H, m), 1.648 (3H, s), 1.329 (3H, s); ¹³C NMR (125 MHz, CDCl₃) δ 153.71, 147.03, 147.00, 139.18, 127.73, 114.91, 111.64, 111.47, 76.60, 43.26, 36.64, 29.64, 25.01, 17.40.